HESI/AAPS Reference Panel for Preclinical Immunogenicity Risk Assessment

Laurent Malherbe Executive Director, ADME Eli Lilly and Company

On behalf of the

AAPS Immunogenicity Risk Assessment and Mitigation Working Group HESI-Immunosafety Method Development Workgroup

EIP 14th Open Symposium, Lisbon, April 2023





- Origins of Reference Panel
- HESI/AAPS Reference Panel Work Proposal
- Preliminary Results

AAPS Immunogenicity Risk Assessment & Mitigation Working Group

- Working group within AAPS Therapeutic Product Immunogenicity Community
- Vision:
 - connect scientists working on preclinical immunogenicity risk assessment and mitigation
 - increase knowledge and understanding of the various risk assessment tools and evaluate their utility
 - provide a forum to discuss challenges facing the field
 - formulate and communicate industry opinions on regulatory challenges related to immunogenicity risk assessment"
- Monthly teleconference (2nd Tuesday of the month)
- Membership:
 - 58 members (US / Europe, Industry/Academia / FDA)

Chair: Sofie Pattijn Former Chair: Laurent Malherbe Vice Chair: Robin Walsh Secretary: Daniel Leventhal

American Association of Pharmaceutical Scientists

Survey outcome on immunogenicity risk assessment tools for biotherapeutics: an insight into consensus on methods, application and utility in drug development

Jochem Gokemeijer,^{1*#} Yi Wen,^{2*} Vibha Jawa,^{3*}, Shibani Mitra-Kaushik⁴, Shan Chung⁵, Alan Goggins⁶, Seema Kumar⁷, Kasper Lamberth⁸, Karen Liao⁹, Jennie Lill⁵, Qui Phung⁵, Robin Walsh², Brian Roberts¹⁰, Michael Swanson¹¹, Inderpal Singh¹², Sophie Tourdot¹³, Mark A. Kroenke¹⁴ Bonita Rup¹⁵, Theresa J. Goletz¹⁶, Swati Gupta¹⁷, Laurent Malherbe², and Sofie Pattijn¹⁸

- 70% of the member companies used some type of predictive assay preclinically
- No alignment on the methods used or data interpretation and criteria used for reporting of predicted risk
- The biggest gaps to a broader implementation of immunogenicity screening were related to:
 - the translatability and predictive value of the preclinical risk assessment data in clinic
 - lack of standardization and benchmarking
 - \succ high variability for these assays.

Are you using in vitro immunogenicity assays



Slide courtesy of Yi Wen, Jochem Gokemeijer, Vibha Jawa and AAPS IRAM work group

Convergence between AAPS and EIP

"While the data generated by these assays and platform is becoming increasingly informative and valuable, the interpretation and translation into an overall immunogenicity risk of a given therapeutic protein remains challenging **due to lack of assay**

standardization and harmonization".

MABS 2022, VOL. 14, NO. 1, e1993522 (17 pages) https://doi.org/10.1080/19420862.2021.1993522

REVIEWS

Taylor & Francis Taylor & Francis Group

OPEN ACCESS

Assay format diversity in pre-clinical immunogenicity risk assessment: Toward a possible harmonization of antigenicity assays

Axel Ducret^a, Chloé Ackaert^b, Juliana Bessa^{a,&}, Campbell Bunce^c, Timothy Hickling^a, Vibha Jawa^d, Mark A. Kroenke^e, Kasper Lamberth^f, Anaïs Manin^c, Hweixian L. Penny^e, Noel Smith^g, Grzegorz Terszowski^h, Sophie Tourdotⁱ, and Sebastian Spindeldreher ¹/₁₀^j

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A sense of Déjà vu : Cytokine Release Assays Development (2014)

- Every pharmaceutical companies developed cytokine release assays to assess the risk of cytokine release syndrome after the cytokine storm experience with TGN 1412 in 2006.
- "It is clear from the HESI survey and follow-up, that there is no standard approach to strategies, assay formats and reporting and interpretation of data, and that pharmaceutical companies, CROs and academic institutions use a variety of approaches."
- A 'one-size-fits-all' approach is not recommended because different biotherapeutics, based on varying mechanisms of actions, may require different approaches.
- However, <u>alignment of technical procedures including</u> <u>positive and negative controls</u>, for frequently used formats and data interpretation and its <u>implementation</u>, as well as regulatory expectations <u>would allow a higher level of harmonization across the</u> <u>scientific community</u>.



HESI-Immuno-safety Technical Committee

 Health and Environmental Sciences Institute (HESI) is a non-profit institution whose mission is to collaboratively identify and help to resolve global health and environmental challenges through the engagement of scientists from academia, government, industry, NGOs, and other strategic partners.

Immuno-safety Technical Committee

- Over 160 active individual active participants across industry, government and academic
- Mission: To identify and address scientific issues related to immune safety and translation to human health risk assessment

– Key Objectives:

- Leverage technical and scientific expertise from academic, regulatory, and industry organizations to advance immuno-safety science
- Contribute to the scientific decision-making processes relative to the development of guidelines and regulations for immune safety testing
- Educate stakeholders in safety science and promote the understanding and appropriate use of immune safety data

HESI-ITC Method Development Workgroup

Method Development Workgroup Co-Leads: Courtni Newsome (BMS), Sandrine Vessillier (NIBSC)

Vision: To share, optimize methodologies used for immune safety testing between companies and research organization to assure appropriate preclinical data for public health safety

WHAT:

- Identify, inform on and assess emerging technologies used by research organizations to address safety liabilities of biologics , small molecules and cell therapy products
- Identify any issues/gaps in the models, assays and data interpretation
- Provide relevant controls to include in the assays

HOW:

- By prioritizing new projects in function of the scientific landscape priorities
- By sharing information on specific methods through survey, SharePoint, teleconferences
- By liaising with other WGs when overlapping interests
- By collaborating with education WG to organize webinar on specific methodology
- By publishing good practices guidelines for the scientific community

WHY:

• To assure a robust use of methods and correct data interpretation



Cytokine Release Assay Reference Panel

"One of the challenges for the development and comparison of CRA performance is the lack of availability of standard positive and negative control mAbs for use in assay qualification.

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- To address this issue, the National Institute for Biological Standards and Control (NIBSC) developed a reference panel of lyophilised mAbs known to induce CRS in the clinic: human anti-CD52, mouse anti-CD3 and human superagonistic (SA) anti-CD28 mAb manufactured according to the respective published sequences of Campath-1H® (alemtuzumab, IgG1), Orthoclone OKT-3® (muromonab, IgG2a) and TGN1412 (theralizumab, IgG4), as well as three isotype matched negative controls (human IgG1, mouse IgG2a and human IgG4, respectively).
- The relative capacity of these control mAbs to stimulate the release of IFN-γ, IL-2, TNF-α and IL-6 in vitro was evaluated in eleven laboratories in an international collaborative study mediated through the HESI Immuno-safety Technical Committee Cytokine Release Assay Working Group."



Development of the first reference antibody panel for qualification and validation of cytokine release assay platforms – Report of an international collaborative study

Sandrine Vessillier^{a,*}, Madeline Fort^b, Lynn O'Donnell^c, Heather Hinton^d, Kimberly Nadwodny^e, Joseph Piccotti^f, Peter Rigsby^a, Karin Staflin^g, Richard Stebbings^h, Divya Mekalaⁱ, Aarron Willinghamⁱ, Babette Wolf^e, participants of the study^{a,b,c,d,e,f,g,b,i,j,k}

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Cytokine Release Assay Panel development Assay diversity

Table 3 CRA performed by the participants.							
Assay type	IncubationTime	Number of participants	Number of donors (all participants)				
PBMC-SP	18–24 h	4	35				
	48 h	3	26				
PBMC-AQ	24 h	1	8				
PBL/HUVEC	24 h	1	8				
WB-AQ	24 h	6	54				
	48 h	1	8				
dWB-AQ	48 h	1	12				
dWB-SP*	48 h	1	15				

PBMC: Peripheral Blood Mononuclear Cells; PBL: peripheral blood leukocytes; HUVEC: human umbilical vein endothelial cells; WB: Whole Blood; dWB: diluted Whole Blood; SP: solid phase; AQ: Aqueous Phase; dWB-SP* corresponds to the bead coated method.



Vessillier, S., et al. (2020). "Development of the first reference antibody panel for qualification and validation of cytokine release assay platforms - Report of an international collaborative study." Cytokine X 2(4): 100042.

Can we develop a reference antibody panel to qualify and validate preclinical immunogenicity assays ?



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What biologics should compose the reference panel?

Desired attributes:

- Homologs to clinically tested therapeutic biologics
- At least one strong positive control that will reproducibly trigger a response in diverse assays
 - > Anti-IL21R antibody (Homolog to ATR-107; 76% ADA+)
- A negative and positive control antibodies targeting the same antigen (mimicking industry preclinical experience)

> Anti-PCSK9 antibody A (Homolog to Evolocumab; <1% ADA+)

> Anti-PCSK9 antibody B (Homolog to Bococizumab; 48% ADA+)

Discussed but not included for cost/feasibility consideration

Immunogenic peptides

What preclinical assays to focus on first?



Reference Panel Development: Activities and Timeline



Pilot study - Characterize the reference panel antibodies using in vitro T cell activation assays

- **Goals:** to assess the activity/immunogenicity of the antibodies produced by Absolute Antibody and to the effect/impact of freeze-drying process on the activity/immunogenicity of the antibodies
- 11 participating laboratories
- Funded by HESI-ITC
- 250-500mg of the 3 reference panel antibodies produced by Absolute Antibodies
- Product Lyophilization at NIBSC and shipment to participating laboratories

Data collection from various T cell assays : Q2-Q3 2023

Data analysis and Manuscript writing : Q3-Q4 2023

Reference Panel Development Status

• 11 Laboratories participating to the pilot study (Abbvie, BMS, Eli Lilly, Epivax, FDA (x2), Genentech, ImmunXperts, Merck, Pfizer, Sanofi)

Assay Type	Incubation Time	Readout	Number of participants
PBMC Assay	7d	IFN-γ	4
PBMC Assay	7d	IL-2	2
PBMC Assay	2d	T cell activation marker	1
PBMC Assay (wo CD8)	7d	Proliferation (CFSE or EdU)	3
DC:T cell Assay	7d	CD4 T Cell Proliferation	2
DC:T cell Assay	7d	IFN-γ	2
DC:T cell Assay	7d	IL-2	1



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Lilly Flow-Based T cell Proliferation Assay



Walsh et al mAb 2020

Previous relevant experience in PBMC assay with internal homologs



Walsh et al mAb 2020

HLA-DR Allotype Expression in cohort used for T cell Proliferation Assessment of the Reference Panel

Donor	DRB1 allele 1	DRB1 allele 2	Ethnicity
1	*07:01	*14:06	Hispanic
2	*07:01	*15:01	Caucasian
3	*07:01	*13:02	African American/Hispanic
4	*04:07	*04:07	Hispanic
5	*08:04	*11:01	African/American
6	*04:02	*12:01	African/American
7	*03:01	*04:01	Caucasian
8	*13:02	*15:03	Hispanic
9	*04:01	*10:01	Caucasian
10	*03:01	*15:01	Hispanic
11	*10:01	*13:05	Caucasian
12	*11:01	*15:03	African/American







T Cell Proliferation Results with Reference Panel Antibodies Representative Pseudocolor Flow Plots



T Cell Proliferation Results with Reference Panel Antibodies across 12 donors





% Positive Donors

Cell Division Index (CDI)

No Impact of Lyophilization on CD4 T cell Proliferative Response to Reference Panel Antibodies



Summary

- There is no alignment on the methods used for assessing the immunogenicity risk.
- One of the challenges for the development and comparison of assay performance is the lack of availability of standard positive and negative control antibodies for use in assay qualification.
- The development of a reference panel of positive and negative control monoclonal antibodies will provide information on the reproducibility, robustness and potential limitations of preclinical assays developed to predict clinical immunogenicity.

Questions?

Interested to join ?

Contacts

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Thank you