Health Agency feedback on immunogenicity assays - Results of an EIP survey

Lydia Michaut
on behalf of the
EIP Immunogenicity Strategy Working Group

EIP Scientific Symposium
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Experience-sharing through survey

32 feedbacks from 11 companies

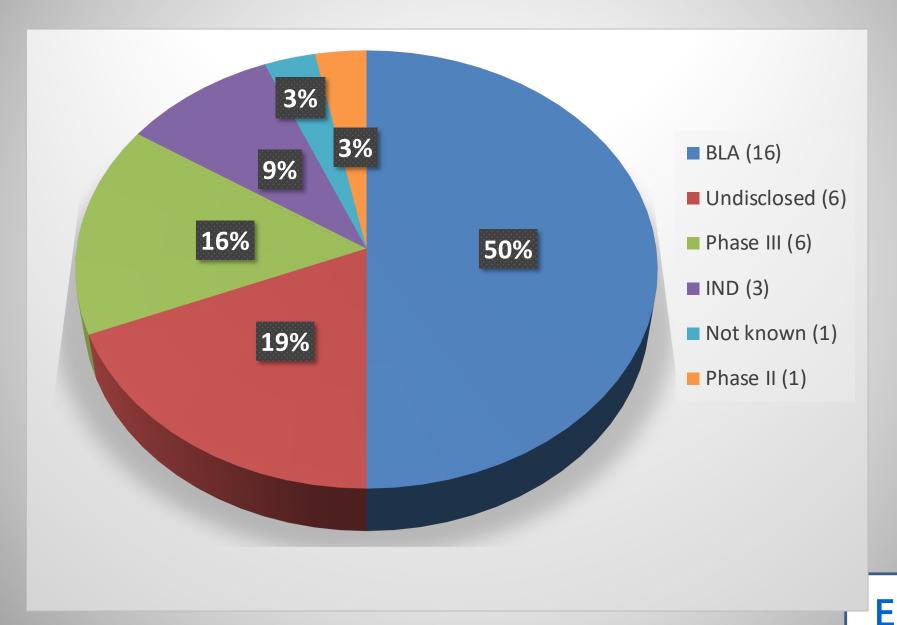
- > 50% participation rate
 (19 companies represented in the EIP Immunogenicity Strategy working group)
- Company names anonymized in the results

«Non-responder» companies (8):

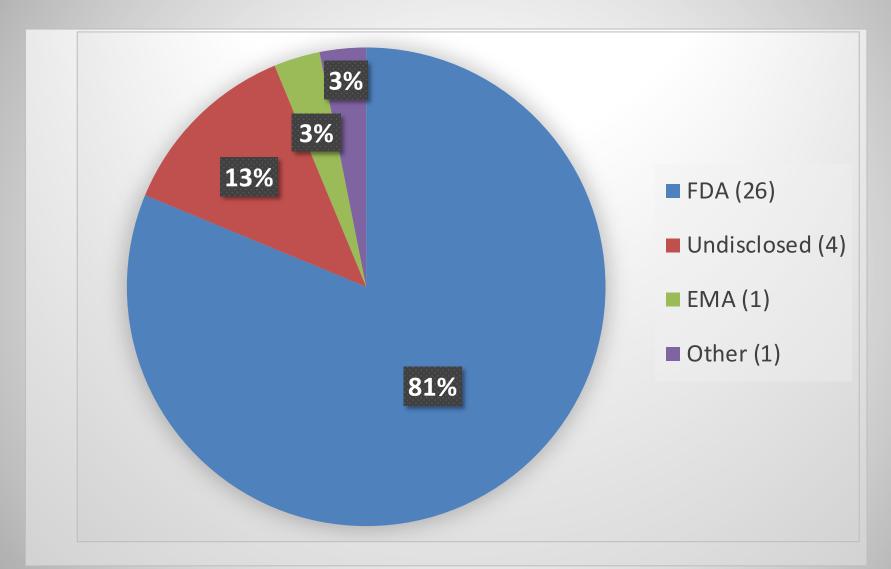
- Audited by / interacted with HA but did not receive any feedback from authorities?
- No audited by / no interaction with HAs?
- Not allowed to share information?



Project development stage (n=32)

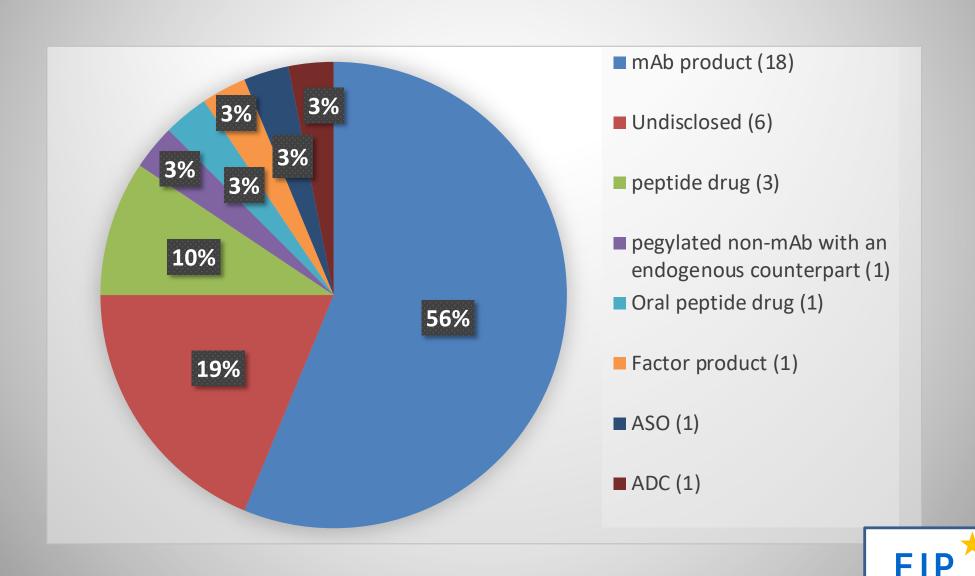


Regulatory agency (n=32)





Drug type



Points of challenge (n= 42)

Assays not meeting guidance requirements: 15

Drug tolerance: 5

NAb assays: 5

Sample collection timepoints: 5

Data to be provided for review - When and What: 4

Project-specific questions: 4

Antibody isotypes: 3

Result reporting: 1



Assays

Request for 1% FPR rather than 0.1% in confirmatory assay (6)

LPC concentration must be established relative to assay sensitivity (4)

Validate selectivity and specificity in hemolyzed serum in the ADA screening and confirmatory assays

Determine inter-assay precision of confirmatory controls

Evaluate prozone effect in the ADA validation

Evaluate selectivity and specificity for the ADA confirmatory assay

Method specificity was not demonstrated for the ADA confirmatory assay in method validation

Drug tolerance

Requested information on drug tolerance levels for ADA levels near the assay sensitivity

Demonstrate that LPC and HPC can be detected in presence of drug levels detected in clinical samples

Justify the acceptability of the Drug Tolerance for the ELISA used in the pivotal studies in context of the expected drug concentrations seen during repeated injection.

Justification of accepted drug tolerance parameter (concentration)

Request for additional drug concentrations to be tested



nAb assays

Format and use

- Of note, the agency generally recommends the use of a cell based neutralizing assay. If you decide to use a competitive ligand binding assay to assess neutralizing antibodies, the IND should be updated with a comparison of the cell based and competitive ligand binding assays
- Agreement on use of competitive ligand binding assay no need for generation of data in cell based Nab assay format
- Rejected proposal to use integrated PK, ADA and target engagement analysis instead of nAb assay

Validation parameters

- Request for 1% FPR in nAb assay
- Request for improved sensitivity and drug tolerance of cell based nAb assay



Sample collection and timepoints

All subjects must be followed/up until reverted to pre-defined titre

Follow up until responses have converted to baseline.

Follow up on positive patients until return to baseline

Sampling time points: D7-10 needed for IgM and IgG at early time points.

Sponsor to collect serum samples earlier in future clinical trials to address the levels of IgM specific ADA



Project-specific questions

Use of a target bead based depletion step in an ADA assay (risk of removal of target-drug-ADA complexes impacting ADA detection)

Demonstrate that the ADA assay can detect immune responses to all components of an ADC (e.g., mAb, ADC, linker-payload)

It is surprising that ADAs have no apparent impact on efficacy despite a clear effect on drug concentration and blood eosinophil level.

Evidence requested that method can detect antibodies in low pH when acid method used



Data to be provided for review - When and What

Agency wish to review all data prior to Ph3 pivotal studies.

Request for "developmental exercises" for all assays.

Data on robustness of the assays requested.

Requirement for adequate storage of all samples under appropriate conditions to allow for additional testing until the agency have reviewed all assays and deem them to be suitable.



Antibody isotypes

Document that assay can detect IgA ADA

Must sensitively detect IgG and IgM

Sensitivity of IgE assay for hypersensitivity samples

Report antibody titers including the MRD



Topic for further discussion

The AAPS Journal (2022) 24: 68 https://doi.org/10.1208/s12248-022-00712-2

COMMENTARY



When to Extend Monitoring of Anti-drug Antibodies for High-risk Biotherapeutics in Clinical Trials: an Opinion from the European Immunogenicity Platform

Gregor P. Lotz¹ · Karin Benstein² · Karien Bloem³ · Harm Buddiger⁴ · Claudio Calonder⁵ · Stefanie Elm⁶ · Elena Fernandez⁷ · Joanne Goodman⁸ · Boris Gorovits⁹ · Joanna Grudzinska-Goebel¹⁰ · Melody Janssen¹¹ · Vibha Jawa¹² · Daniel Kramer² · Linlin Luo¹³ · Mantas Malisauskas¹⁴ · Lydia Michaut¹⁵ · Martin Schäfer¹ · Sebastian Spindeldreher¹⁵ · Martin Ullmann¹⁶ · Karin Nana Weldingh¹⁷ · Arno Kromminga^{18,19} · Veerle Snoeck²⁰

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General schematic ADA sample collection in clinical studies

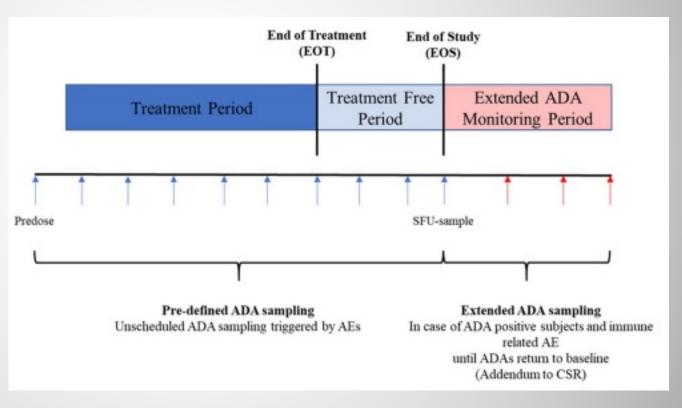
ADA sampling during:

- drug treatment period
- treatment-free period-> until end of study.

Extended ADA Monitoring Period:

In case of a high risk for serious consequences from ADAs, regulators expect to extend sampling for ADA positive subjects beyond EOS until ADA levels return to baseline

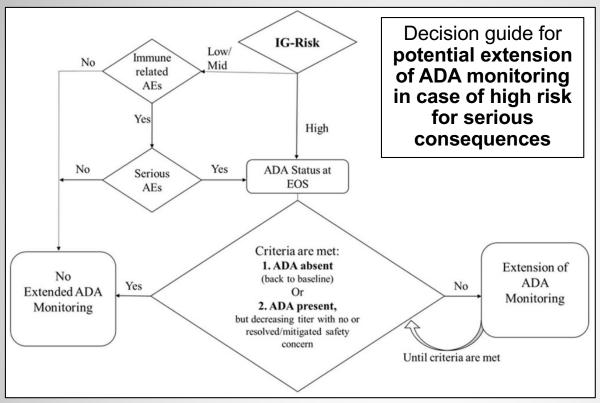
Arrows indicate collection time-points



ADA=Anti-drug antibodies; SFU=Safety Follow-Up; CSR=Clinical Study Report AE=adverse event; EOS=End of Study



EIP Opinion



At EOS, when ADAs, are either absent or still detectable but ADA titers are decreasing, and safety concerns are either absent or resolved/mitigated then no extension of ADA monitoring is required.

ADA=Anti-drug antibodies; SFU=Safety Follow-Up; AE=adverse event; EOS=End of Study

ADA extended monitoring should be based on evaluation of ADA data in the context of corresponding clinical signals:

If the clinical data shows that safety consequences are minor, mitigated, or resolved, then further ADA monitoring may not be required despite potentially detectable ADAs above baseline.

Thank you

EIP Immunogenicity Strategy Working Group co-leaders

Veerle Snoeck
Jo Goodman
Arno Kromminga
Lydia Michaut

Member company representives:

UCB Merck

MedImmune / AZ Novartis

BioAgilytix NovoNordisk

Abzena Pfizer

Amgen Roche

Boehringer Ingelheim Sandoz

Covance Sanofi

Genmab Sanquin

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